

Selenium nanoparticles and their utilization in scaffolds

Prof. RNDr. Vojtech Adam, Ph.D.
Mgr. Dagmar Hegerova, Ph.D.
Mendel University in Brno

Content

2

1. Nanoparticles – silver, selenium
 - *Staphylococcus aureus*; Bacterial infections
 - Antibiotic resistance
2. Product SelenBact
3. Nanoparticles in scaffolds

Staphylococcus aureus

3

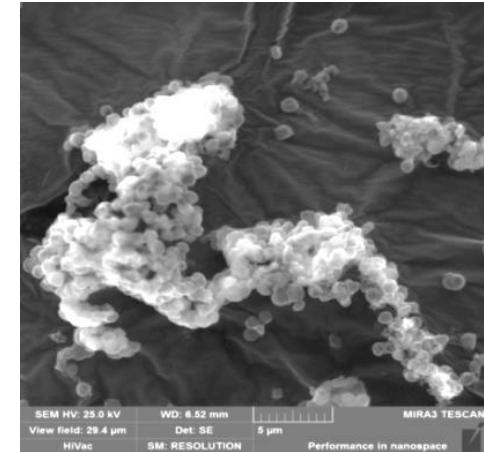
- G⁺, facultative anaerobic coccal bacterium of the genus *Staphylococcus* ($\varnothing 1 \mu\text{m}$)
- In 1/3 of the population naturally presented in the human respiratory tract, on the skin and mucous membranes



Colonies of *S. aureus*



Colonies of *S. aureus* on Petri dish

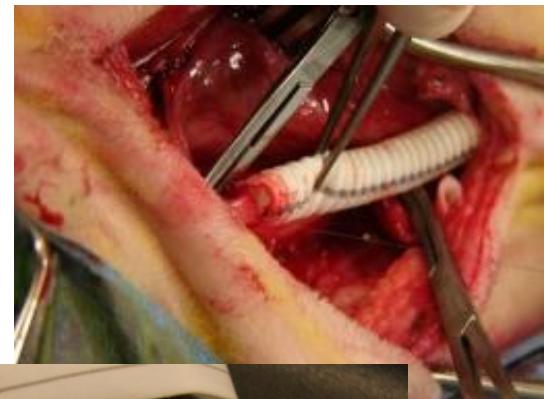


S. aureus under electron microscopy

Staphylococcus aureus

4

- Not always pathogenic, but often a serious agent of the bacterial infections, emerging after transplantation of implants (e.g. vascular grafts etc.)
- Disease-associated strains often promote infections by producing potent protein toxins, and expressing cell-surface proteins that bind and inactivate antibodies



Bacterial infections: treatment and prevention

5

- Localized infections – surgically
- 90 % of staphylococci are resistant to penicillin → use of penicillin, which inhibits beta-lactamase
- Macrolides, cephalosporins, aminoglycosides, tetracyclines etc.
- Prevention – vaccines to stimulate specific immunity
- Antibiotic-resistant strains (e.g. MRSA) = worldwide problem in clinical medicine

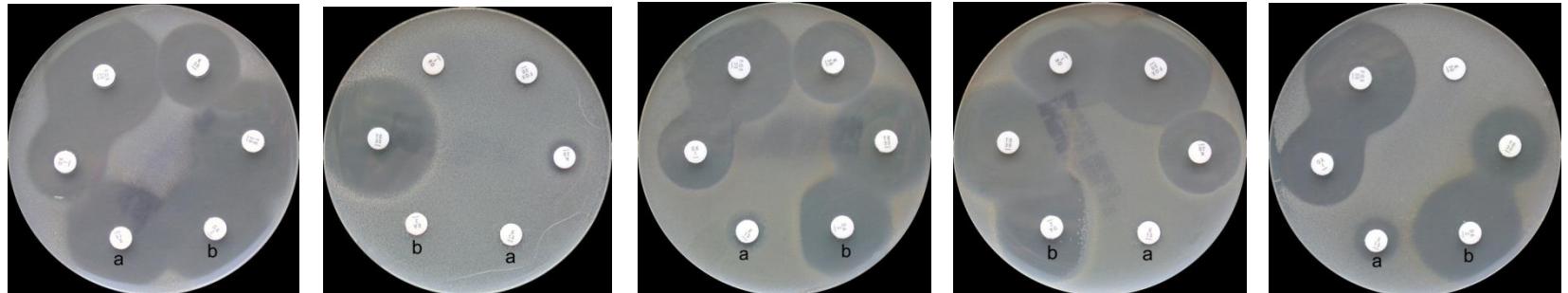


Photo Credit: Gregory Moran, M.D.

Resistance of microorganisms

6

- Biological resistance = resistance x environment effects,
- Resistance genes code genetic information, which can be changed by environmental factors.



Characterization of nanoparticles

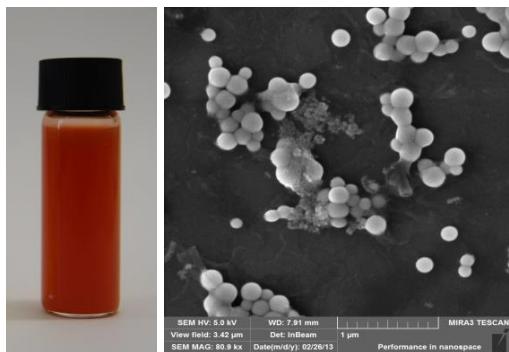
7

- Silver phosphate nanoparticles (SPNPs)



- (200-300 nm) square or spherical character

- Selenium nanoparticles (SeNPs)



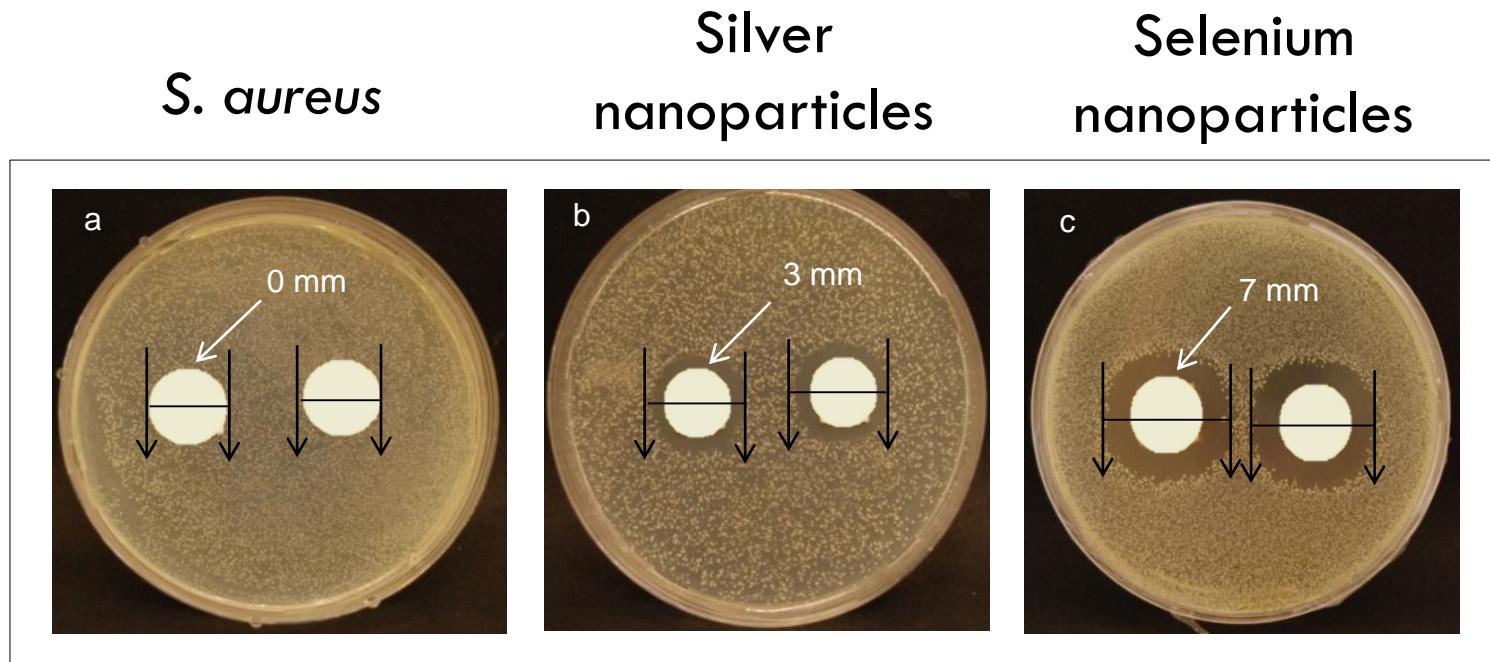
- (50-100 nm) small spherical particles

Chudobova et al., Comparison of the effects of silver phosphate and selenium nanoparticles on Staphylococcus aureus growth reveals potential for selenium particles to prevent infection FEMS Microl. Letters, 351(2), 195-201

Antimicrobial effect of nanoparticles

8

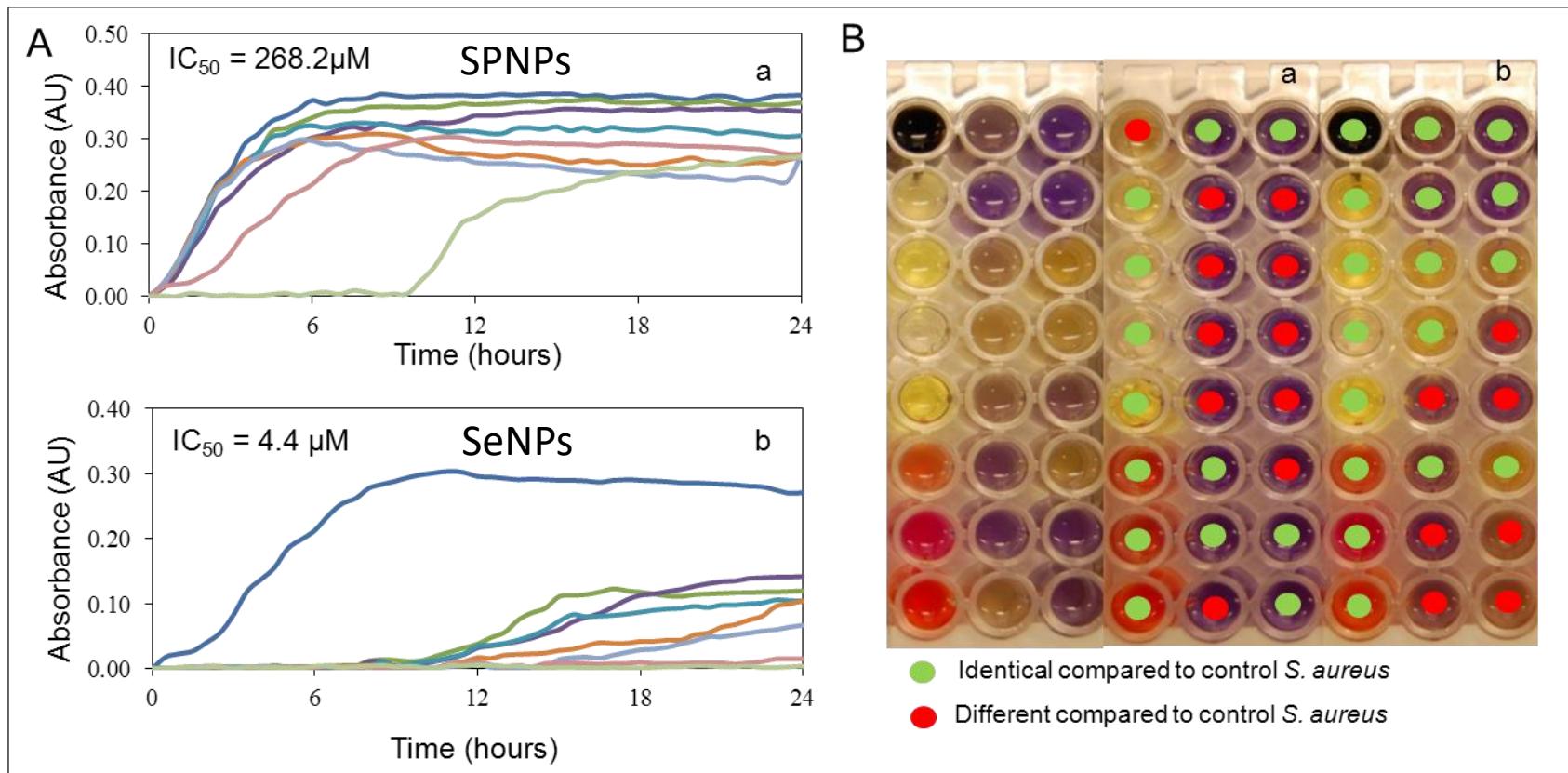
- Growth inhibition zones in bacterial culture after application of $300 \mu\text{M}$ concentration of nanoparticles
- Significant growth inhibition after application of smaller-sized SeNPs (even twice larger zones - 7 mm) were observed in comparison with larger SPNPs (3 mm zones)



Antimicrobial effect of nanoparticles

9

- (A) Spectrophotometric analysis of the growth of *S. aureus* bacterial culture with SPNPs (a) or SeNPs (b) in concentrations of ● 0, ● 10, ● 25, ● 50, ● 75, ● 150, ● 225 and ● 300 μM and (B) colorimetric study of changes in biochemical parameters after application of 300 μM concentration of nanoparticles (B).



Product SelenBact

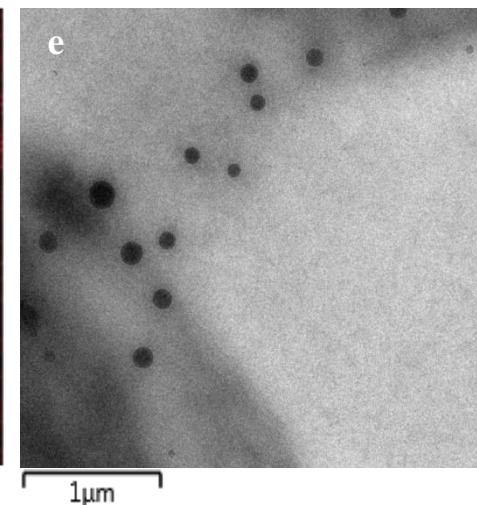
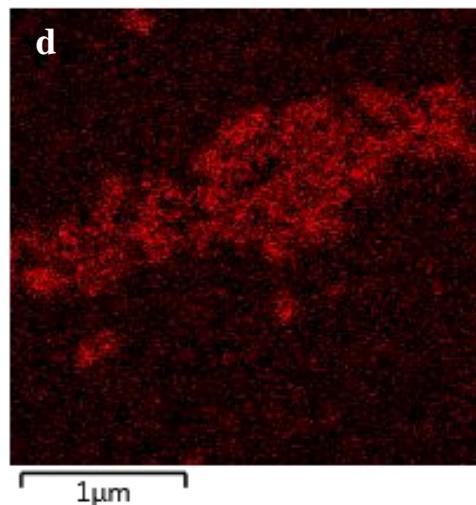
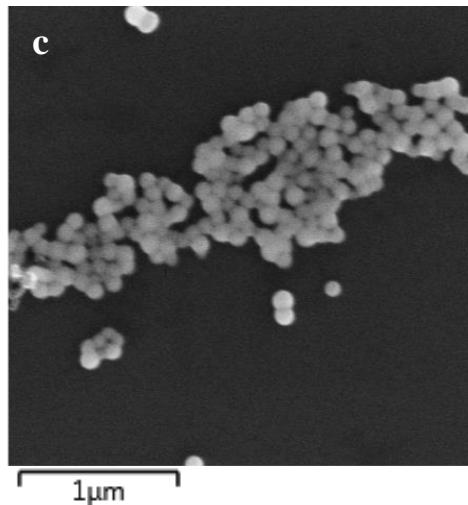
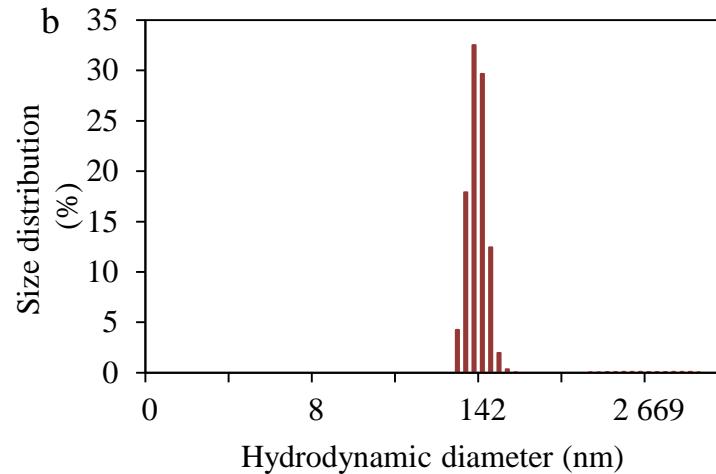
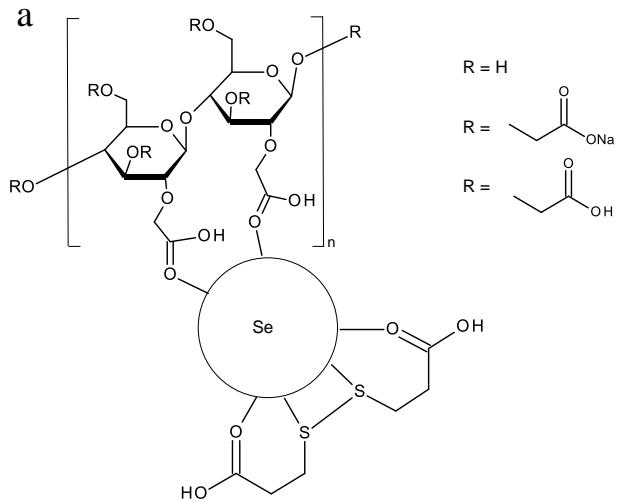
11

- MedicProgress Ltd.
- Characteristics:
 - pH 6.8 – 7.2
 - Content of Se 158 ppm \pm 10 %
(selenium content in solution was characterized by AAS)
 - Size 50 – 100 nm
 - Viscosity 1500 – 4500 mPa.s
 - Color – orange



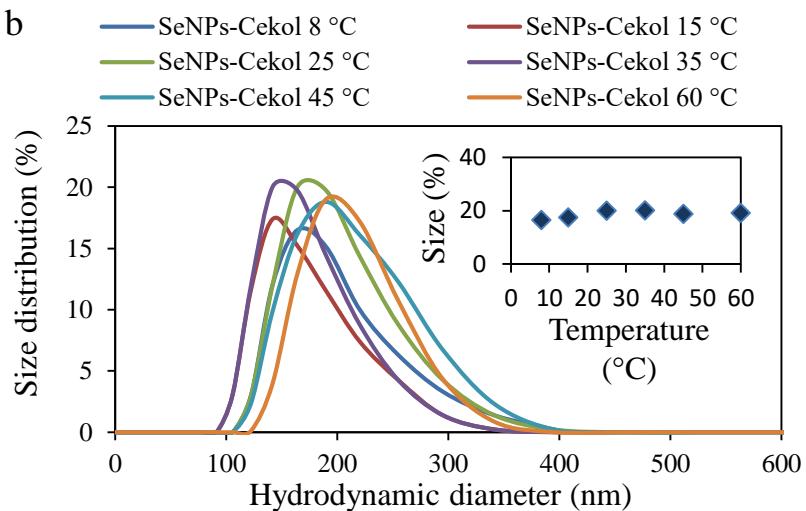
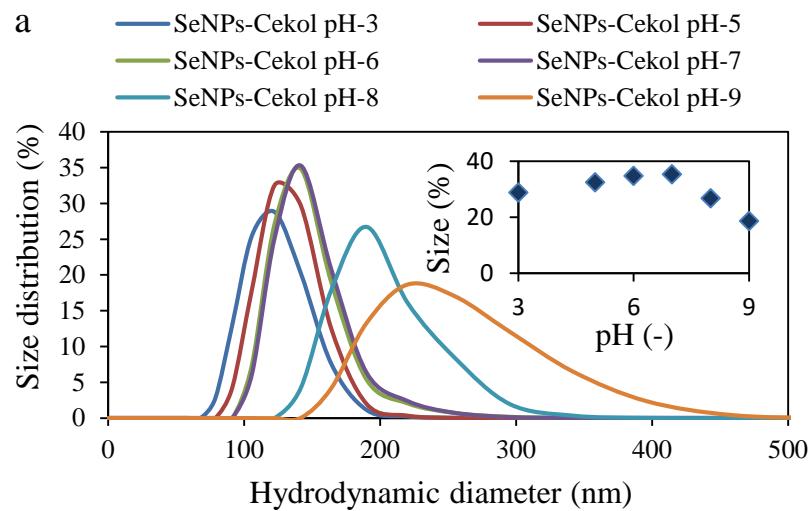
Chemical structure of SelenBact

12



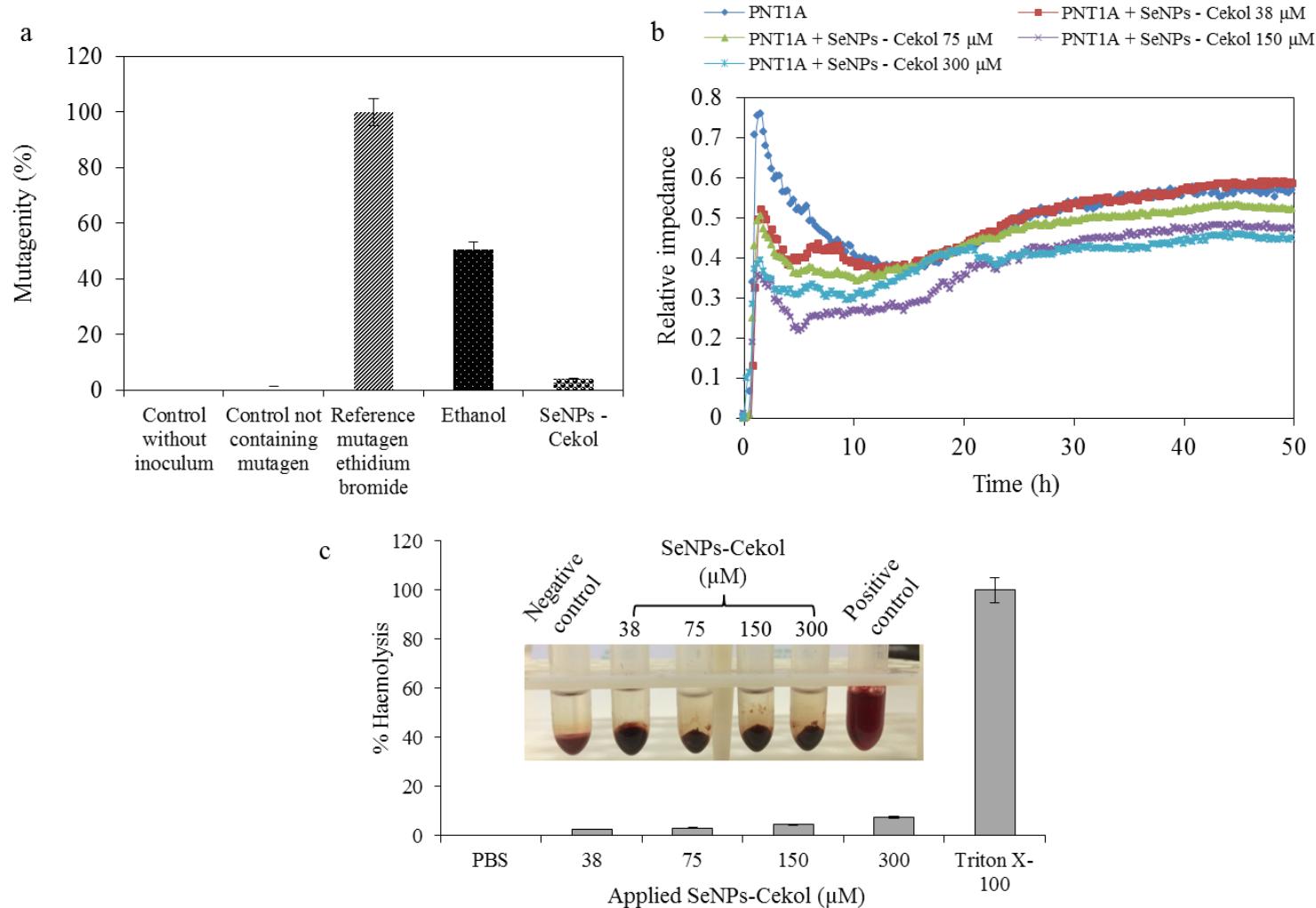
Determination of SelenBact stability

13



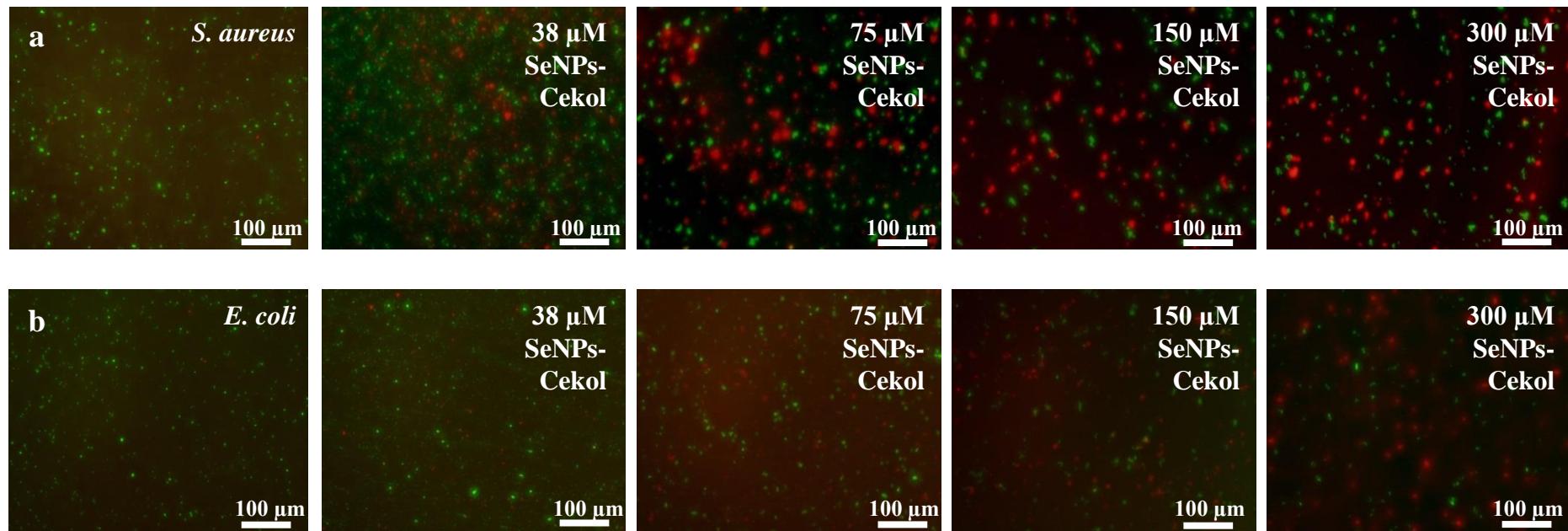
Testing of toxicity and mutagenicity of SelenBact on eukaryotic cells

14



Fluorescence microscopy of bacteria treated with SelenBact

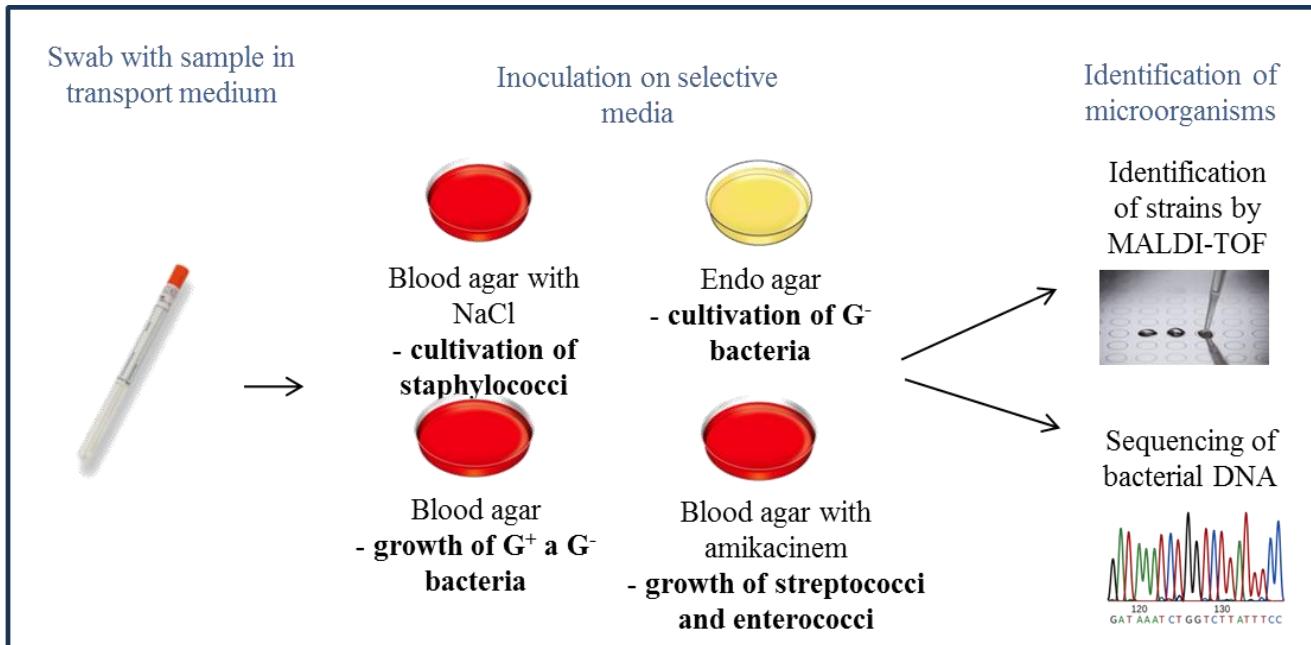
15



Bacterial isolates identification by MALDI

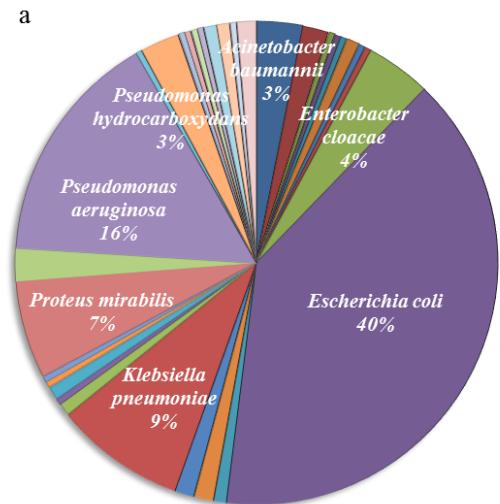
16

- Cooperation with Trauma Hospital in Brno
- Collection of swabs (300 patients) from surface wounds infected by bacterial specimens
- Isolation of these strains
- Identification by different methods (microbiology, PCR, sequencing, MS)



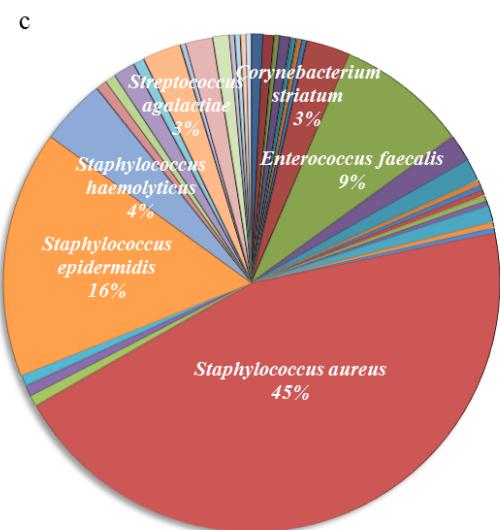
Bacterial isolates identification by MALDI

17



b

Specie of gram-negative bacteria	Occurence (%)	Specie of gram-negative bacteria	Occurence (%)
<i>Acinetobacter baumannii</i>	3.1	<i>Neisseria perflava</i>	0.9
<i>Acinetobacter genomospecies</i>	1.7	<i>Neisseria weaveri</i>	0.4
<i>Acinetobacter Iwoffii</i>	0.4	<i>Pantoea agglomerans</i>	0.4
<i>Aeromonas caviae</i>	0.4	<i>Proteus mirabilis</i>	6.6
<i>Aeromonas media</i>	0.4	<i>Proteus vulgaris</i>	2.2
<i>Alcaligenes faecalis</i>	0.9	<i>Pseudomonas aeruginosa</i>	15.7
<i>Arthrobacter parietis</i>	0.4	<i>Pseudomonas alcaligenes</i>	0.4
<i>Citrobacter freundii</i>	0.4	<i>Pseudomonas hydrocarboxydans</i>	2.6
<i>Enterobacter cloacae</i>	4.4	<i>Pseudomonas mendocina</i>	0.4
<i>Escherichia coli</i>	39.7	<i>Pseudomonas nitroreducens</i>	0.4
<i>Escherichia fergusonii</i>	0.9	<i>Pseudomonas oryzihabitans</i>	0.4
<i>Hafnia alvei</i>	1.3	<i>Pseudomonas syringae</i>	0.4
<i>Klebsiella oxytoca</i>	1.3	<i>Raoultella ornithinolytica</i>	0.9
<i>Klebsiella pneumoniae</i>	8.7	<i>Serratia marcescens</i>	0.9
<i>Morganella morganii</i>	0.9	<i>Serratia weitica</i>	0.4
<i>Neisseria gonorrhoeae</i>	0.4	<i>Stenotrophomonas maltophilia</i>	1.3

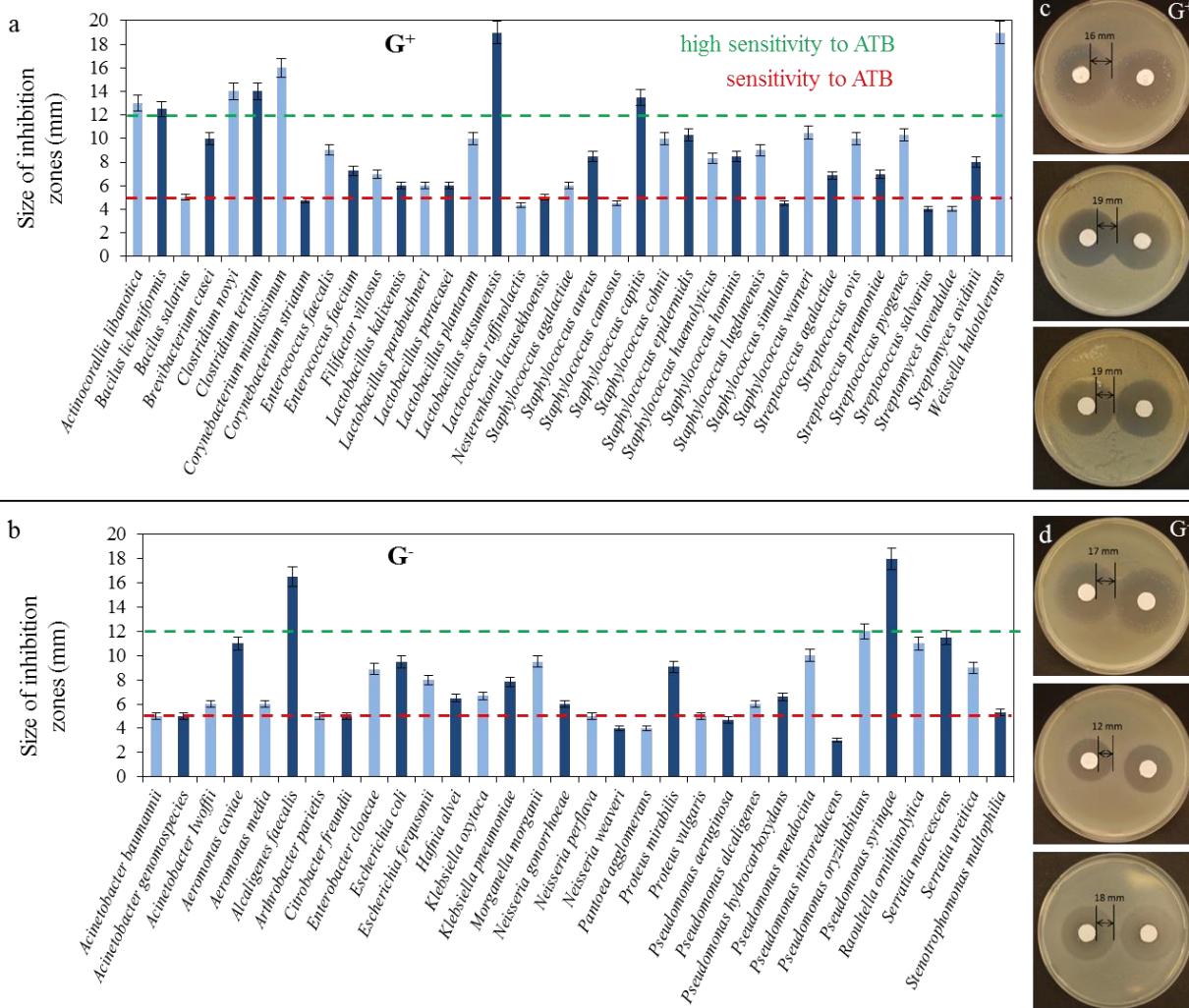


d

Specie of gram-positive bacteria	Occurence (%)	Specie of gram-positive bacteria	Occurence (%)
<i>Actinocorallia libanotica</i>	0.7	<i>Staphylococcus aureus</i>	45.0
<i>Bacillus licheniformis</i>	0.7	<i>Staphylococcus camosus</i>	0.7
<i>Bacillus salarius</i>	0.4	<i>Staphylococcus capitis</i>	0.7
<i>Brevibacterium casei</i>	0.7	<i>Staphylococcus cohnii</i>	0.7
<i>Clostridium novyi</i>	0.4	<i>Staphylococcus epidermidis</i>	16.1
<i>Clostridium teritum</i>	0.4	<i>Staphylococcus haemolyticus</i>	4.3
<i>Corynebacterium minutissimum</i>	0.4	<i>Staphylococcus hominis</i>	0.7
<i>Corynebacterium striatum</i>	2.9	<i>Staphylococcus lugdunensis</i>	0.7
<i>Enterococcus faecalis</i>	8.6	<i>Staphylococcus simulans</i>	1.4
<i>Enterococcus faecium</i>	1.8	<i>Staphylococcus warneri</i>	0.7
<i>Filifactor villosum</i>	1.4	<i>Streptococcus agalactiae</i>	2.5
<i>Lactobacillus kalmensis</i>	0.4	<i>Streptococcus ovis</i>	0.4
<i>Lactobacillus parabuchneri</i>	0.4	<i>Streptococcus pneumoniae</i>	1.8
<i>Lactobacillus paracasei</i>	0.4	<i>Streptococcus pyogenes</i>	1.1
<i>Lactobacillus plantarum</i>	0.4	<i>Streptococcus salvarius</i>	0.4
<i>Lactobacillus satsumensis</i>	0.4	<i>Streptomyces lavendulae</i>	0.4
<i>Lactococcus raffinolactis</i>	1.1	<i>Streptomyces avidinii</i>	0.4
<i>Nesterenkonia lacusekhoensis</i>	0.4	<i>Weissella halotolerans</i>	0.4
<i>Staphylococcus agalactiae</i>	0.4		

Effects of SelenBact on bacterial isolates

18



Selenium nanoparticles in scaffolds

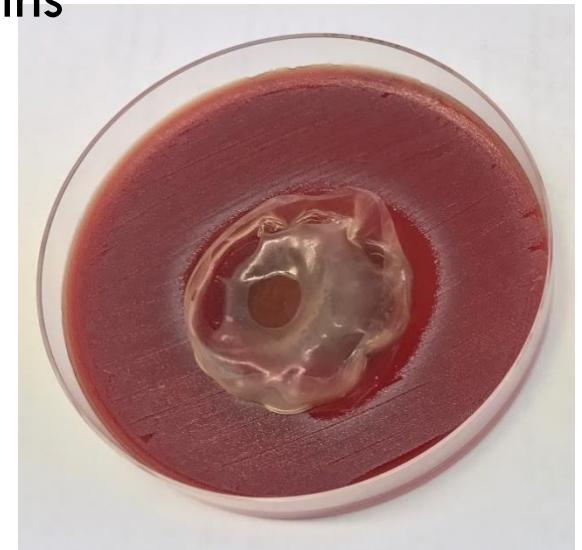
19

- Advanced nanomaterials – potential in treatment of surface bacterial infections
- Attachment to the surface of cover material
 - Long-term sterility
 - Elimination and reduction the risk of spreading bacterial infection
 - Acceleration of tissue regeneration
 - Reduction the cost of treating infections

Selenium nanoparticles in scaffolds

20

- Physical and chemical fixation by cold plasma
- Promising results in terms of excellent transparency
- Non-adherent specimens – newly created tissue
- Ensuring the inhibition of the pathogens
 - In the contact area
 - Across the spectrum of bacterial strains



Antimicrobiality of SeNPs on scaffolds

21

- Surface treated with plasma
- Samples
 - ref. + SeNPs 0.2%
 - ref + SeNPs 0.2% + 1s plasma
 - 1s plasma + SeNPs 0.2%
 - 5s plasma + SeNPs 0.2%
 - 1s plasma + SeNPs 0.2% + 1s plasma
 - 1s plasma
 - 5s plasma
 - Control (pure fabric)

Size of inhibition zones

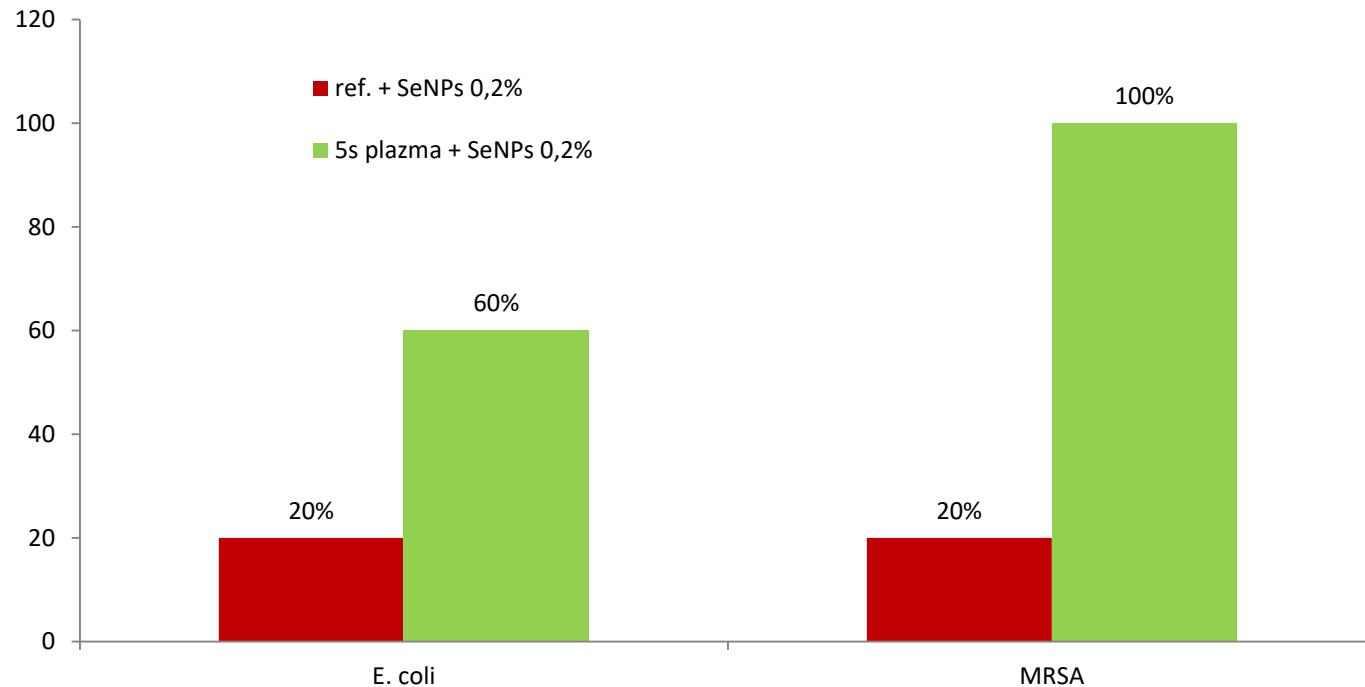
22

Fabrics	<i>E. coli</i>	MRSA
1. ref. + SeNPs 0.2%	1	1
2. ref + SeNPs 0.2% + 1s plasma	0	1
3. 1s plasma + SeNPs 0.2%	2	2
4. 5s plasma + SeNPs 0.2%	3	5
5. 1s plasma + SeNPs 0.2% + 1s plazma	2	4
6. 1s plasma	1	0
7. 5s plasma	1	0
8. Control (pure fabric)	0	0

* Most suitable to reduce bacterial growth is fabric 5s plasma + SeNPs 0.2%

Comparison of effects of SeNPs and SeNPs with plasma

23



Acknowledgment

24

Thank you for your attention

